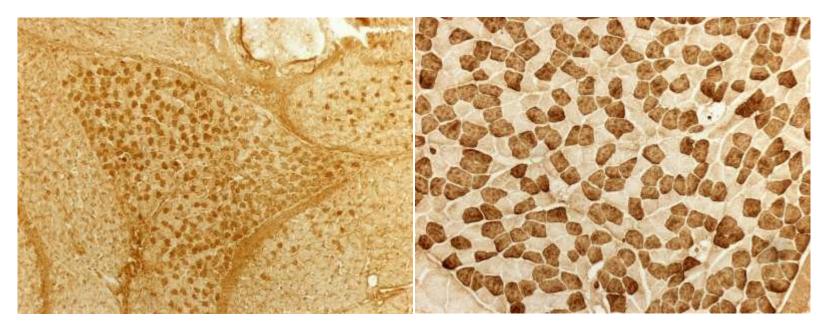
Exercise 1: Estimate the degree of MATURATION HYPERTROPHY that occurs in mouse SOLEUS muscle fibres between birth and adulthood

Images taken at same magnification – low power

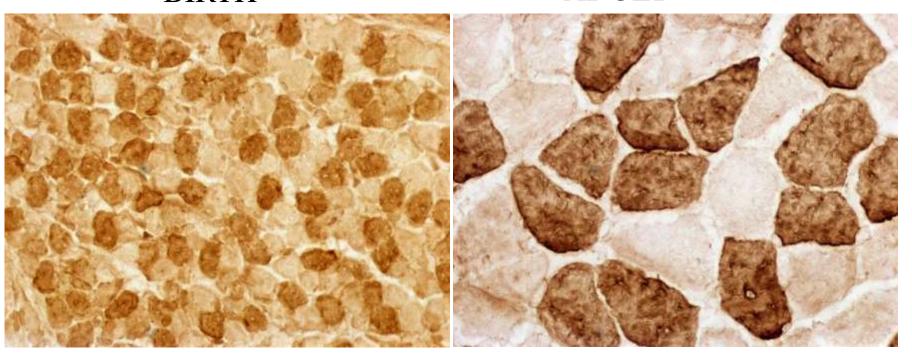
BIRTH ADULT



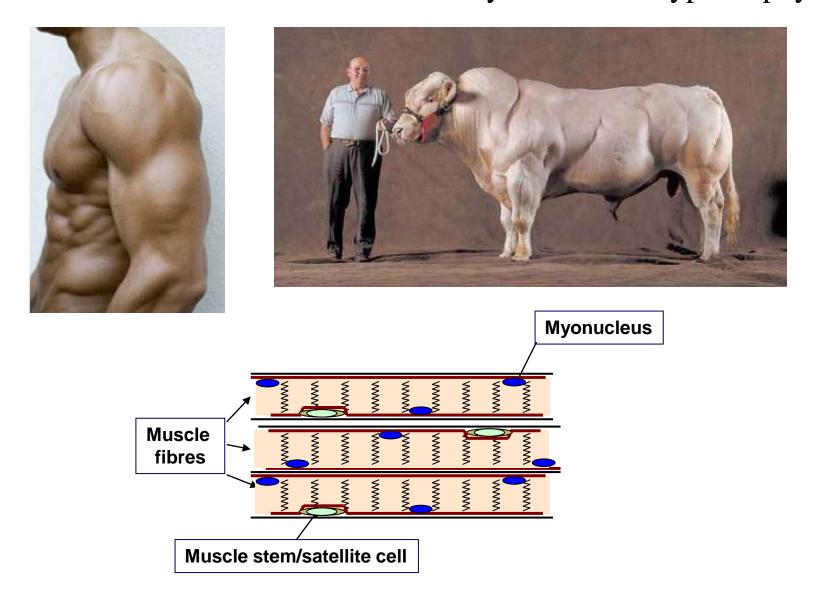
Exercise 1: Estimate the degree of MATURATION HYPERTROPHY that occurs in mouse SOLEUS muscle fibres between birth and adulthood

Images taken at same magnification – HIGH power

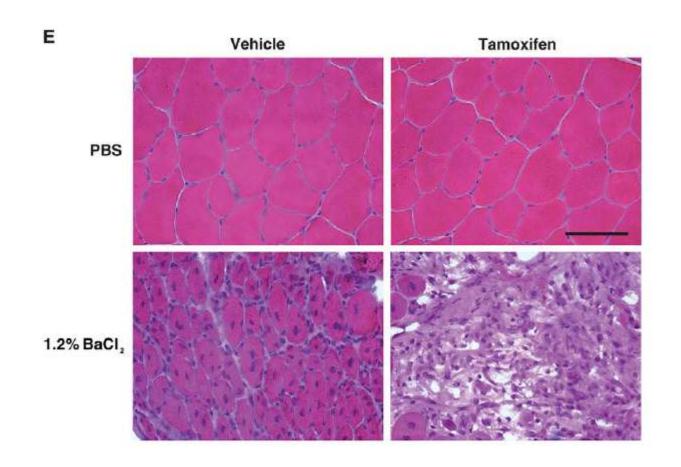
BIRTH ADULT



Exercise 2: Are satellite cells necessary for muscle hypertrophy?

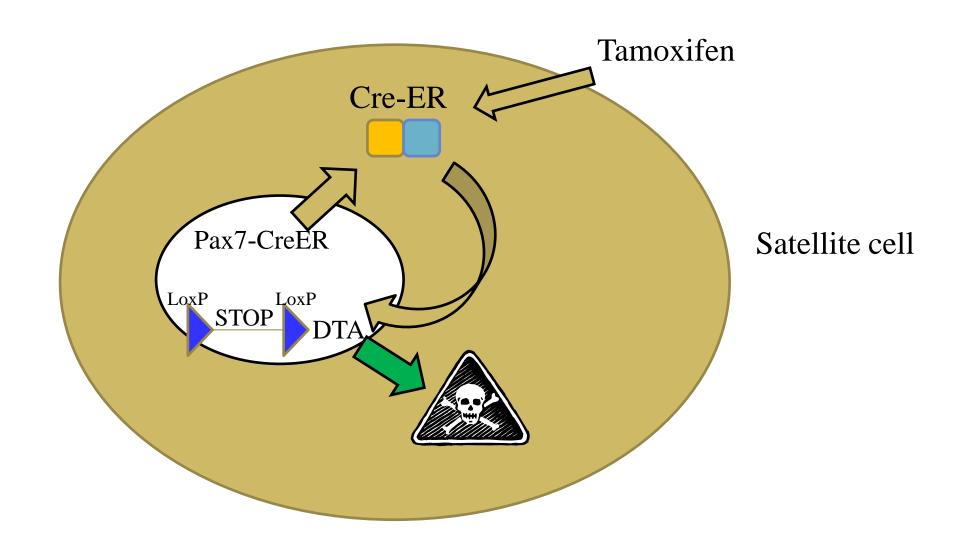


Pax7^{CreER/CreER} mice crossed with Floxed DTA mice Treated with Tamoxifen to induce the Cre expression

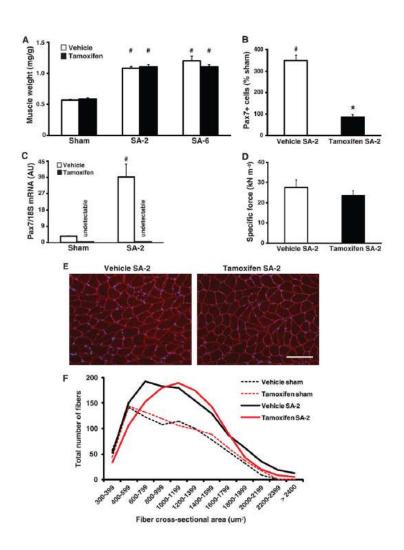


Barium Chloride BaCl2 causes muscle damage

CRE-ER: Cre DNA recombinase fused to the estrogen receptor **Floxed-DTA**: Diptheria toxin A gene flanked by LoxP sites, which are recombined by the Cre protein



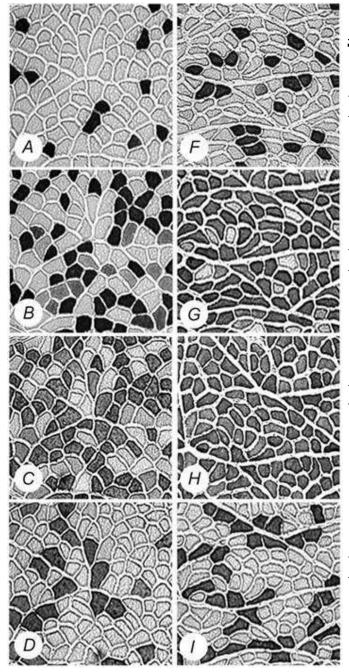
Synergist ablation (SA): removing nearby muscles to induce hypertrophy experimentally in the Pax7-DTA mice



Exercise 2: read the results section of the McCarthy et al. 2011 paper

Discuss

Exercise 3: Experimental induction of muscle fibre type change Using Chronic low frequency stimulation (CLFS)



Antibody

MyHC I

MyHC IIa

All MyHC but not IIx

MyHC IIb

Exercise 3: Chronic Low Frequency Stimulation Experiment – Rat left common peroneal nerve stimulated 21 days with CLFS. Transverse sections of Tibialis Anterior muscle stained with Antibodies against the different MyHC protein isoforms (dark coloured fibres)

A,B,C,D – Control no CLFS F,G,H,I – CLFS for 21 days

Discuss findings

The Journal of Physiology

A publication of The Physiological Society

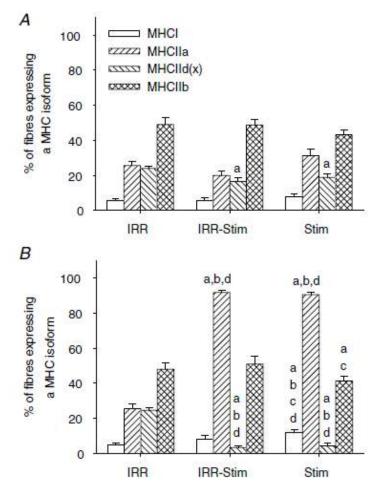
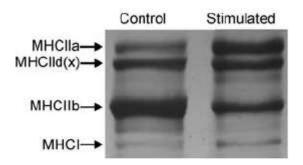


Figure 6
The percentage of fibres expressing a particular MHC isoform in contralateral control (A) and stimulated/sham rat tibialis anterior muscles (B). Statistical symbols as in Fig. 2.

Measuring fibre type conversion using immunohistochemistry on muscle sections



Measuring fibre conversion using protein gels

Figure 8

Example of the electrophoretic method used to quantify MHC isoform composition of rat tibialis anterior muscles. Control (IRR-control) and stimulated (Stim) are shown.

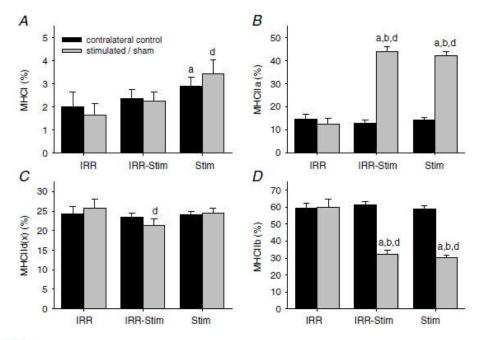


Figure 9
Percentage of MHCI (A), MHCIIa (B), MHCIId(x) (C) and MHCIIb (D) distribution as determined by densitometric evaluation of triplicate gels. Statistical symbols as in Fig. 2.